

FORM PTO-1390 (Modified)
(REV 11-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

204742US0PCT

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

09/787893

INTERNATIONAL APPLICATION NO.

PCT/SE99/01741

INTERNATIONAL FILING DATE

01 October 1999

PRIORITY DATE CLAIMED

01 October 1998 (earliest)

TITLE OF INVENTION

REDUCTION OF OXIDATIVE STRESS FACTORS

APPLICANT(S) FOR DO/EO/US

Marek NARUSZEWICZ

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ Certificate of Mailing by Express Mail
20. ☒ Other items or information:

**Request for Consideration of Documents Cited in International Search Report
Notice of Priority**

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.53) <div style="font-size: 24pt; font-weight: bold; text-align: center;">09/787893</div>		INTERNATIONAL APPLICATION NO. <div style="font-weight: bold; text-align: center;">PCT/SE99/01741</div>		ATTORNEY'S DOCKET NUMBER <div style="font-weight: bold; text-align: center;">204742US0PCT</div>	
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21. The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

☒ Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO **\$1,000.00**

☐ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO **\$860.00**

☐ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO **\$710.00**

☐ International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) **\$690.00**

☐ International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) **\$100.00**

ENTER APPROPRIATE BASIC FEE AMOUNT =

\$1,000.00

CALCULATIONS PTO USE ONLY

Surcharge of **\$130.00** for furnishing the oath or declaration later than ☐ 20 ☒ 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).

\$130.00

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	14 - 20 =	0	x \$18.00		\$0.00
Independent claims	2 - 3 =	0	x \$80.00		\$0.00
Multiple Dependent Claims (check if applicable).				<input type="checkbox"/>	\$0.00
TOTAL OF ABOVE CALCULATIONS =					\$1,130.00
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable).					\$0.00
SUBTOTAL =					\$1,130.00
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).					\$0.00
TOTAL NATIONAL FEE =					\$1,130.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).					\$0.00
TOTAL FEES ENCLOSED =					\$1,130.00
					Amount to be: refunded \$
					charged \$

☒ A check in the amount of **\$1,130.00** to cover the above fees is enclosed.

☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees.

A duplicate copy of this sheet is enclosed.

☒ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. **15-0030** A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

END ALL CORRESPONDENCE TO:

22850

Surinder Sachar
Registration No. 34,423

Surinder Sachar

SIGNATURE

Norman F. Oblon

NAME

24,618

REGISTRATION NUMBER

March 28 2001

DATE

09/787893

204742US0 PCT

JCO8 Rec'd PCT/PTO 28 MAR 2001

IN THE UNITED STATES PATENT & TRADEMARK OFFICE

IN RE APPLICATION OF: :
MAREK NARUSZEWICZ : ATTN: NEW APPLICATION DIVISION
SERIAL NO: NEW APPLICATION :
(Based on PCT/SE99/01741)
FILED: HEREWITH :
FOR: REDUCTION OF OXIDATIVE :
STRESS FACTORS

PRELIMINARY AMENDMENT

ASSISTANT COMMISSIONER FOR PATENTS
WASHINGTON, D.C. 20231

SIR:

Prior to examination on the merits, please amend the above-identified application as follows:

IN THE CLAIMS

Please amend the claims as shown in the attached marked-up copy to read as follows:

- 3. (Amended) Use according to claim 1, wherein the bacterial strain is a *Lactobacillus plantarum* strain.
4. (Amended) Use according to claim 1 of *Lactobacillus plantarum* 299v, deposition number DSM 9843.
5. (Amended) Use according to claim 1 for the reduction of IL-6 in blood.
6. (Amended) Use according to claim 1 for the reduction of the level of reactive oxygen species in blood.
7. (Amended) Use according to claim 1 for the reduction of the adhesion of

monocytes to endothelial cells.

8. (Amended) Use according to claim 1 for the manufacture of a medicament for the prophylaxis and/or treatment of chronic inflammatory diseases.--

Please add the following claims:

--11. (New) A method of reducing the levels of oxidative stress factors in the blood of a mammal comprising administering to a mammal in need thereof an effective amount of a bacterial strain which gives rise to a significant increase in the faecal concentration of propionic acid to said mammal.

12. (New) The method according to claim 11, wherein said bacterial strain is a strain of *Lactobacillus* or *Propionibacterium*.

13. (New) The method according to claim 11, wherein said bacterial strain is *Lactobacillus plantarum*.

14. (New) The method according to claim 11, wherein said bacterial strain is *Lactobacillus plantarum* 299v, deposition number DSM 9843.--

REMARKS

Claims 1-14 are active in the present application. The claims are amended to remove multiple dependencies. Claims 11-14 find support in Claims 1-10 and the specification. No new matter is added. An action on the merits and allowance the claims is solicited.

Respectfully submitted,

OBLON, SPIVAK, McCLELLAND,
MAIER & NEUSTADT, P.C.



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Marked-Up Copy

Serial No: _____

Amendment Filed on: _____

IN THE CLAIMS

Please amend the claims as follows:

- 3. (Amended) Use according to claim 1 [or 2], wherein the bacterial strain is a *Lactobacillus plantarum* strain.
4. (Amended) Use according to [any of claims 1 to 3] claim 1 of *Lactobacillus plantarum* 299v, deposition number DSM 9843.
5. (Amended) Use according to [any of claims 1-4] claim 1 for the reduction of IL-6 in blood.
6. (Amended) Use according to [any of claims 1-4] claim 1 for the reduction of the level of reactive oxygen species in blood.
7. (Amended) Use according to [any of claims 1-4] claim 1 for the reduction of the adhesion of monocytes to endothelial cells.
8. (Amended) Use according to [any of claims 1-4] claim 1 for the manufacture of a medicament for the prophylaxis and/or treatment of chronic inflammatory diseases.--

REDUCTION OF OXIDATIVE STRESS FACTORS

The present invention refers to the use of one or more bacterial strains to reduce the level of oxidative stress factors in mammals including man.

BACKGROUND OF THE INVENTION

Oxidative stress factors is the common term for a number of molecules primarily causing prooxidant conditions in the body. Said factors, such as IL-1, IL-6, ROS (reactive oxygen species), IL-8, 8-isoprostaglandin, VCAM (vascular cell adhesion molecule) and ICAM (intracellular adhesion molecule) are present at an elevated level in proinflammatory and inflammatory states.

The oxidative stress factors can also be designated inflammatory markers, which expression, however, in addition comprises so called secondary molecules, which are initiated by the oxidative stress factors, for example acute phase proteins.

ROS, as well as the other oxidative stress factors, are produced by the monocytes and lymphocytes. Normal production of the specific level serve to maintain homeostasis in the body.

An elevated level of oxidative stress factors is typical for acute and chronic inflammation. In chronic inflammation there is a risk of an increased ageing process, atherosclerosis and cancer. Chronic inflammatory states are for instance induced by heavy smoking or by chronic infections with viruses and bacteria. Another group of chronic inflammatory diseases comprises autoimmune conditions such as rheumatic diseases and psoriasis.

Chronic inflammation patients are today treated with antibiotics, high doses of vitamins or other drugs. The use of antibiotics should for several reason be avoided and the use of drugs is mostly associated with different unwanted side-effects. Rheumatic diseases are for instance treated with the drug ibuprofen which is effective but expensive and gives gastric side-effects. For psoriasis patients there is no real medication.

PRIOR ART

EP 0 649 603 A1 in the name of Otsuka Pharmaceutical Co.,

Ltd. refers to an antioxidant food comprising a fermentation product of a food containing a manganese-containing natural material, such as tea leaves, by means of a microbe such as *Lactobacillus plantarum*. The reason for choosing said bacterium is said to be the catalase activity as well as superoxide dismutase-like activity thereof, which brings about an antioxidant activity in the body, but only in the presence of manganese.

Nenonen, MT., et al., British Journal of Rheumatology, March 1998, 37(3) p 274-81, describes the effect of uncooked lactobacilli-rich vegan food in rheumatoid patients. Half of the patients experienced adverse effects such as nausea and diarrhoea and therefore stopped the experiment, but the other half experienced a subjective release of symptoms. It is speculated that a daily consumption of lactobacilli might have a positive effect also on objective measures of rheumatoid arthritis.

US 4,314,995 in the name of Seikenkai refers to a process for treating a patient having an infection or an inflammation caused by an infectious disease, which comprises administering at least one *Lactobacillus* which has different nutritional requirements compared to known strains of *Lactobacillus*, that is which has the ability to grow on a special low nutrition culture medium. Five different strains of *Lactobacillus* are mentioned as being of the invention.

International Journal of Food Microbiology, 42 (1998) 29-38, discloses a significant increase in the total faecal concentration of the short-chain fatty acids (SCFA) acetic acid and propionic acid after 3 weeks of intake of 400 ml/d of a rose-hip drink containing oats fermented with the probiotic *Lactobacillus plantarum*. This increase, which is said to be independent from the basal diet, can either be explained by a production of SCFA by the administered probiotic strain or by said strain stimulating or suppressing other SCFA producing bacteria in the colon.

DESCRIPTION OF THE INVENTION

The present invention refers to the use of a bacterial

strain giving rise to increased amounts of propionic acid in the gut for the manufacture of a medicament for reduction of the level of oxidative stress in mammals including man.

The invention especially refers to the use of a bacterial strain giving rise to increased amounts of propionic acid in the gut for the manufacture of a medicament for reduction of the level of IL-6, ROS, and the adhesion of monocytes to endothelial cells in mammals including man.

A high level of oxidative stress factors, such as the cytokines interleukin 1 and interleukin 6, ROS, and the adhesion of monocytes to endothelial cells is a characteristic of proinflammatory and inflammatory states.

The bacterial strain giving rise to increased amounts of propionic acid in the gut is preferably a strain of *Lactobacillus* or *Propionibacterium*.

According to a preferred embodiment of the invention the bacterial strain is a *Lactobacillus plantarum* strain. A preferred strain of *Lactobacillus plantarum* is the strain *Lactobacillus plantarum* 299v, which has been deposited at the DSM, Deutsche Sammlung von Mikroorganismen von Zellkulturen GmbH, Braunschweig, Germany, under the accession number DSM 9843.

The invention also refers to the use of a bacterial strain giving rise to increased amounts of propionic acid in the gut for the manufacture of a medicament for the prophylaxis and/or treatment of chronic inflammatory diseases. Chronic inflammatory or proinflammatory diseases which can be treated according to the invention can be induced by different bacteria, such as *Chlamydia pneumoniae* and *Helicobacter pylori*, or toxic substances, such as nicotine. It has for instance been demonstrated that the number of antibodies against *Helicobacter pylori* was reduced after one month consumption of Pro Viva, a rose-hip drink containing oats fermented with *Lactobacillus plantarum* DSM 9843 (5×10^7 cfu/ml) in an amount of 400 ml/d.

A preferred use according to the invention is for the prophylaxis and/or treatment of autoimmune diseases, such as rheumatic diseases, and psoriasis.

DESCRIPTION OF THE DRAWINGS

Figure 1 shows the reduction of the adhesion of monocytes to endothelial cells after treatment with fermented oatmeal gruel;

Figure 2 shows the reduction of the adhesion of monocytes to stimulated endothelial cells after treatment with fermented oatmeal gruel;

Figure 3 shows the generation of ROS in monocytes before and after treatment with fermented oatmeal gruel.

EXPERIMENTAL

The purpose of the following experiments was to determine the effect of the administration of a probiotic bacterial strain giving rise to increased concentrations of propionic acid in the gut on the level of oxidative stress factors in blood in subjects having an elevated level of said factors. Each person was given 25 ml/d for either 3 or 6 weeks of a concentrated oatmeal gruel fermented with *Lactobacillus plantarum* DSM 9843 (containing 1×10^9 cfu/ml).

Methods

Peripheral mononuclear cells (PBMC)

PBMC were isolated from heparinized blood by density-gradient centrifugation. The blood was diluted (1:1) in PBS. 25 ml of diluted blood was immediately layered over 15 ml Ficoll-Paque and centrifuged (1900 rpm, 40 min, 22°C). The mixed mononuclear cell band was removed by aspiration and washed with PBS. Isolated PBMC was counted and divided into two parts. One part was used to determine intracellular ROS production, second one was used to adhesion assay. The mononuclear cell preparation consists of approximately 30 % monocytes and 70 % lymphocytes.

Endothelial cell isolation and culture

Human umbilical vein endothelial cells (HUVEC) were obtained from umbilical cords by collagenase digestion, as described by Jaffe, E.A., et al., Culture of human endothelial cells derived from umbilical veins, J. Clin Invest, 1973; 52: 2745-2756. In brief, vein of umbilical cords were perfused with PBS to remove blood cells, filled with 0.1 % collagenase (Ia type) and left for 10 min at 37°C. The endothelial cell (EC)

suspension was supplemented with FBS, fetal bovine serum, and centrifuged at 1200 rpm for 10 min. EC were cultured in M-199 medium under humidified atmosphere at 5 % CO₂ at 37°C. The medium also contained 100 U/ml penicillin, 100 µg/ml streptomycin, 2.5 µg/ml fungizone, 20 mM HEPES, 20 % FBS and 50 µg/ml endothelial cell growth supplement (ECGS). HUVEC were cultured in gelatin-coated 25 cm² flasks. The medium was replaced every 2 days until the cells attained confluence (3-5 days). HUVEC purity was assessed by a "cobble-stone" morphology typical for quiescent EC and factor VIII staining. Confluent HUVEC were detached by 0.01% trypsin/EDTA antagonized by FBS.

Test 1. Effect on ROS production

In this study the effect of the administration of a concentrated oatmeal gruel fermented with *Lactobacillus plantarum* DSM 9843 to a group of six healthy volunteers with high levels of reactive oxygen species, ROS, due to heavy smoking, was evaluated. The medium age was 32 years and the body mass index 26.6 kg/m². Each person was given 25 ml/d for a period of 3 weeks.

Blood was collected from the six individuals. Peripheral blood mononuclear cells (PBMC) were separated by Gradisol L density gradient centrifugation as described above.

The measurement of cell oxidation is based on reactive oxygen species (ROS) mediated conversion of nonfluorescent 2,7'-dichlorofluorescein (DCFH), loaded into cells as 2,7'-dichlorofluorescein diacetate, into fluorescent DCF with increased fluorescence emission reflecting enhanced oxidative stress. Freshly isolated PBMC, consisting of approximately 30 % monocytes and 70 % lymphocytes, were resuspended in phosphate buffered saline (PBS) followed by incubation with 20 µM 2',7'-dichlorofluorescein at 37°C for 30 min in the dark. The cells were then washed with PBS. The relative fluorescence intensity of the fluorophore 2',7'-dichlorofluorescein, which is formed by peroxide oxidation of its nonfluorescent precursor, was detected with cytofluorimetric assay (FACScan, Becton Dickinson). During flow cytometric analysis monocytes and lymphocytes were gated on the basis of forward scatter (FCS) and side scatter (SCC). The

results are expressed as mean fluorescence intensity in the following Table 1.

Table 1.

Reactive oxygen species (ROS) production by monocytes and lymphocytes isolated from subjects before and after treatment with *L. plantarum* 299v

Individual	Monocytes		Lymphocytes	
	Before treatment	After treatment	Before treatment	After treatment
1	220	120	86	81
2	254	369	150	134
3	562	249	217	169
4	425	201	172	135
5	343	204	188	129
6	338	224	133	106

The above data show that the production of ROS was reduced in all individuals except number 2, which is a no-responder.

Test 2. Effect of ibuprofen on ROS production, a comparative study

In order to investigate the mechanisms behind the reduction of ROS by ibuprofen, a propionic acid derivative and a well-known antiinflammatory drug, ibuprofen was administered in an amount of 500 mg/d to the same group of six healthy volunteers as participated in the study described in Test 1 for 3 weeks. Preliminary data indicate that the monocytes and the lymphocytes respond in exactly the same way as to the *Lactobacillus plantarum* 299v in the above study.

Test 3. Effect on production of ROS

The study described in Test 1 was repeated on a group of 10 healthy volunteers (heavy smokers), which were given the concentrated oatmeal gruel for 6 weeks. The results, expressed

expressed as mean fluorescence intensity before and after administration of the concentrated oatmeal gruel are given in Figure 3, which illustrates the generation of ROS in normal resting monocytes before and after administration of *Lactobacillus pl. 299v*.

Test 4. Adhesion assay

PBMC from the same group of 10 healthy volunteers as in Test 3 was also tested as to adhesion in the following way.

For adhesion studies HUVEC of the passage 3 were cultured in gelatin-coated 24-well plates. When confluent monolayers were formed, medium was changed to medium 199 containing antibiotics, 20% FBS and 20 mM HEPES without ECGS 18 hours before experiments. Some cells were pretreated with TNF α (500 u/ml) for 18 hours. Freshly isolated PBMC were resuspended in medium 199 with 20 mM HEPES to a concentration $3 - 9 \times 10^5$ /ml. HUVEC were washed with PBS before addition of PBMC (0,5 ml per well) and coincubated for 30 min. The PBMC suspension was withdrawn. HUVEC were washed with PBS and the wells were fixed with formalin, stained with May-Grunwald and Giemsa solution according to Pappenheim and the number of adherent monocytes were counted in 10 separate areas. Data are expressed as percentage of monocyte added. All experiments were performed in duplicate.

The results are given in Figure 1, illustrating the adhesion of monocytes to non-stimulated HUVEC before and after administration of *Lactobacillus pl. 299v*, and in Figure 2, illustrating the adhesion of normal resting monocytes to TNF- α -stimulated HUVEC before and after administration of *Lactobacillus pl. 299v*.

Test 5. Effect on interleukin 6 (IL-6)

PBMC from the same group of 10 healthy volunteers as in Test 3 was also tested as to the effect of the concentrated oatmeal gruel on the level of IL-6.

The level of IL-6 was measured by the h-Interleukin-6 ELISA kit from Boehringer Mannheim, which is a photometric enzyme immunoassay for the quantitative in vitro determination of human interleukin-6 (hIL-6) in streptavidin-coated microtiter plates. The results are given in the following Table 2.

Table 2.

Subject No.	IL-6 (pg/ml)	IL-6 (pg/ml)
1	10	9
2	11	5
3	7	11
4	13	10
5	14	8
6	5	5
7	10	5
8	7	8
9	15	9
10	9	6

Test 6. Propionic acid induced reduction of oxidative stress

Human monocyte macrophage cells were isolated as described above and 6 million cells were preincubated in 2 ml of RPMI-1640 medium (10 % fetal calf serum, 50 mg/ml of penicillin/streptomycin, 2 mM glutamine) in dishes having a diameter of 35 mm. 10 μ M propionic acid was then added and the cells were incubated at 37°C for 4 h. The medium was subsequently removed and replaced with fresh medium and the cells were stimulated with LPS (lipopolysaccharide), 1.0 ng/ml medium, in order to provoke prooxidant stress. After 24 h the medium was removed and concentrated 10 times by evaporation. Oxidative stress with and without (control cells) propionic acid was evaluated using the 8-Isoprostane EIA kit from Cayman Chemical.

The reduction of oxidative stress expressed as the mean value from 3 experiments of the decrease in the 8-isoprostane level was 47 %.

Test 7. Reduction of oxidative stress factors in psoriasis patients

6 patients with relapsing psoriasis volunteered to participate in the study. The patients were untreated, that is

without medication for at least 6 weeks before the start of the administration of the concentrated oatmeal gruel fermented with *Lactobacillus plantarum* DSM 9843 (containing 1×10^9 cfu/ml). Each patient was given 25 ml/d for 6 weeks.

Peripheral mononuclear cells (PBMC) were isolated from said patients before and immediately after the end of the administration of *Lactobacillus*, as described above. The following oxidative stress factors were measured: ROS, before and after stimulation with PMA, phorbol miristate acetate, (100 ng/ml), which stimulates the cells to oxidative stress; adhesion of monocytes to endothelial cells; and IL-6. The results, obtained by the same methods as described above, are given in the following Table 3.

Table 3.

Reduction of oxidative stress factors in psoriasis patients before and after administration of *Lactobacillus pl.* 299v

Patient No.		ROS (mfi*)		Adherence (%)	IL-6 (pg/ml)
		before PMA	after PMA		
1	before	774	4209	22	25
	after	781	1524	13	14
2	before	1039	5442	23	18
	after	1033	3803	11	9
3	before	660	1797	26	32
	after	1175	4087	11	19
4	before	695	3772	-	29
	after	818	1516	-	17
5	before	840	3708	30	25
	after	1348	5905	20	19
6	before	564	4019	9,3	23
	after	1034	5520	13	21

*mfi = mean fluorescence intensity

In a clinical evaluation of the patients after 6 weeks treatment 5 of the 6 patients showed a significant decrease in itching. In addition the affected area of the skin had been divided into smaller areas; the decrease in area varied from 10 to 27 %.

CONCLUSION

It is suggested that propionate produced in the large gut by colonic microbial fermentation may have an antiinflammatory effect. It is therefore believed that bacterial strains which give rise to increased amounts of propionic acid in the gut will decrease the proinflammatory state connected with different chronic inflammatory disorders in the body.

CLAIMS

1. Use of a bacterial strain giving rise to a significant increase in the faecal concentration of propionic acid for the manufacture of a medicament for reduction of the level of oxidative stress factors in blood in mammals including man.
2. Use according to claim 1, wherein the bacterial strain is a strain of *Lactobacillus* or *Propionibacterium*.
3. Use according to claim 1 or 2, wherein the bacterial strain is a *Lactobacillus plantarum* strain.
4. Use according to any of claims 1 to 3 of *Lactobacillus plantarum* 299v, deposition number DSM 9843.
5. Use according to any of claims 1-4 for the reduction of IL-6 in blood.
6. Use according to any of claims 1-4 for the reduction of the level of reactive oxygen species in blood.
7. Use according to any of claims 1-4 for the reduction of the adhesion of monocytes to endothelial cells.
8. Use according to any of claims 1 to 4 for the manufacture of a medicament for the prophylaxis and/or treatment of chronic inflammatory diseases.
9. Use according to claim 8, for the prophylaxis and/or treatment of rheumatic diseases.
10. Use according to claim 8, for the prophylaxis and/or treatment of psoriasis.

FIG. 1

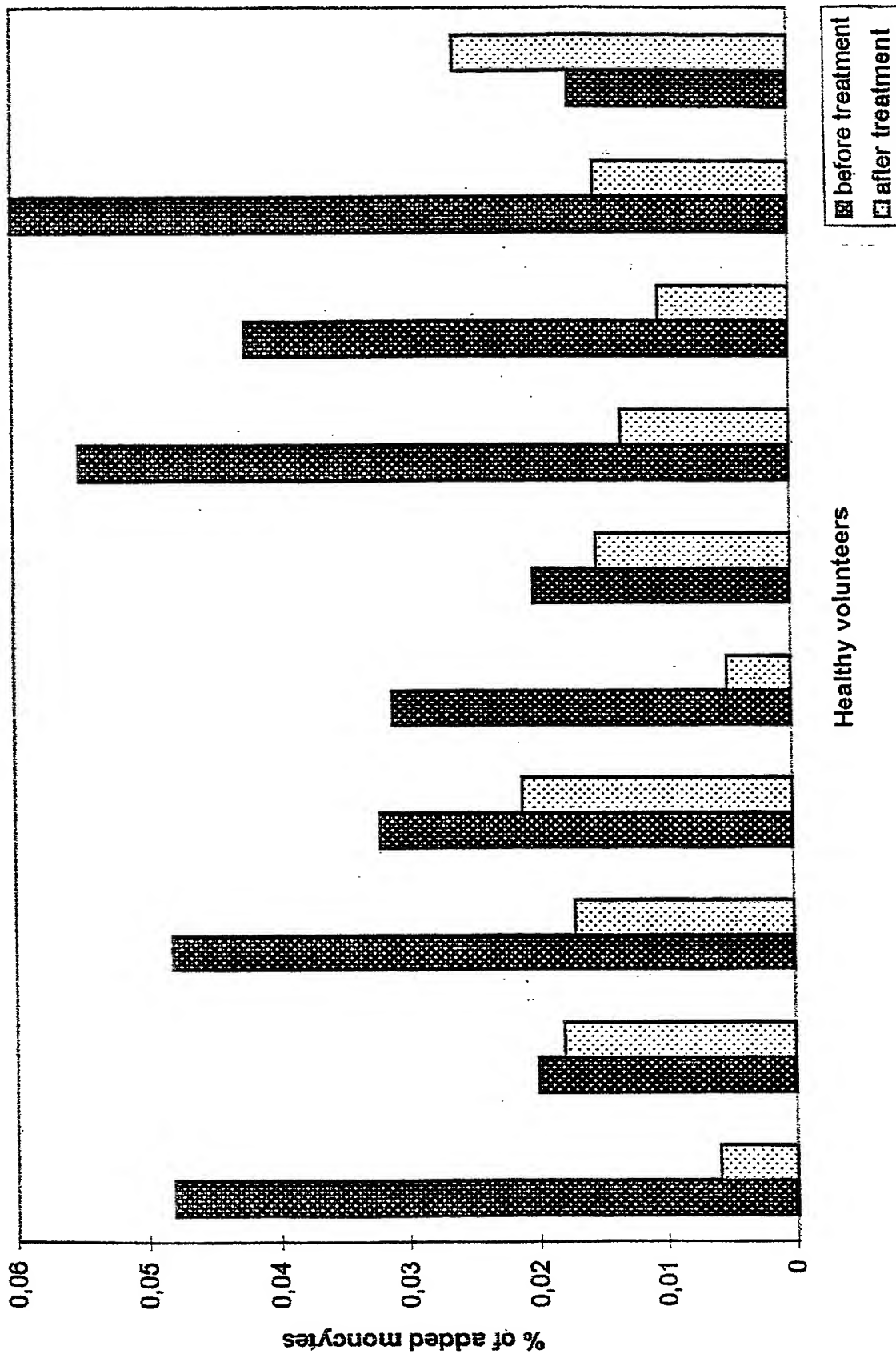


FIG. 2

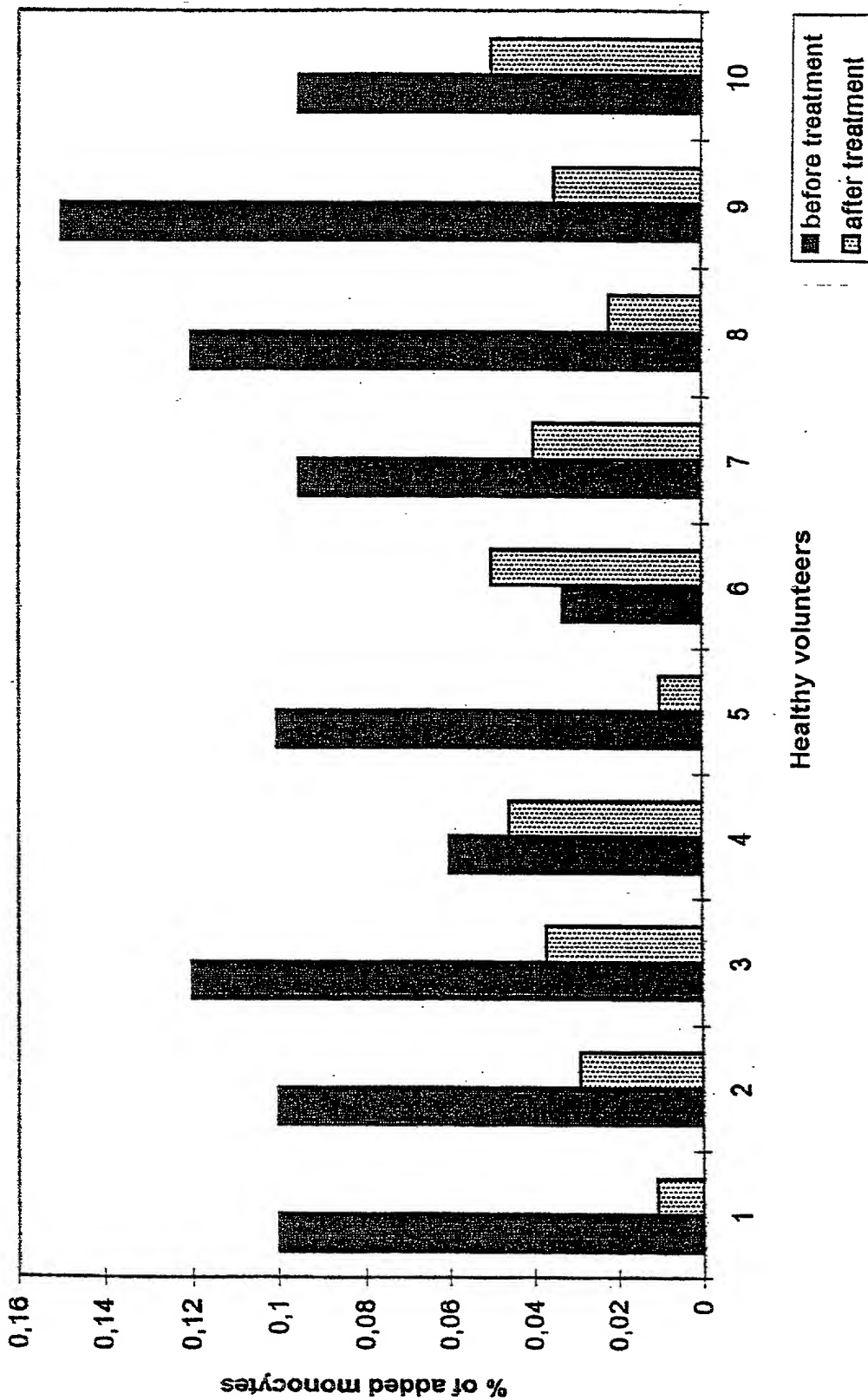
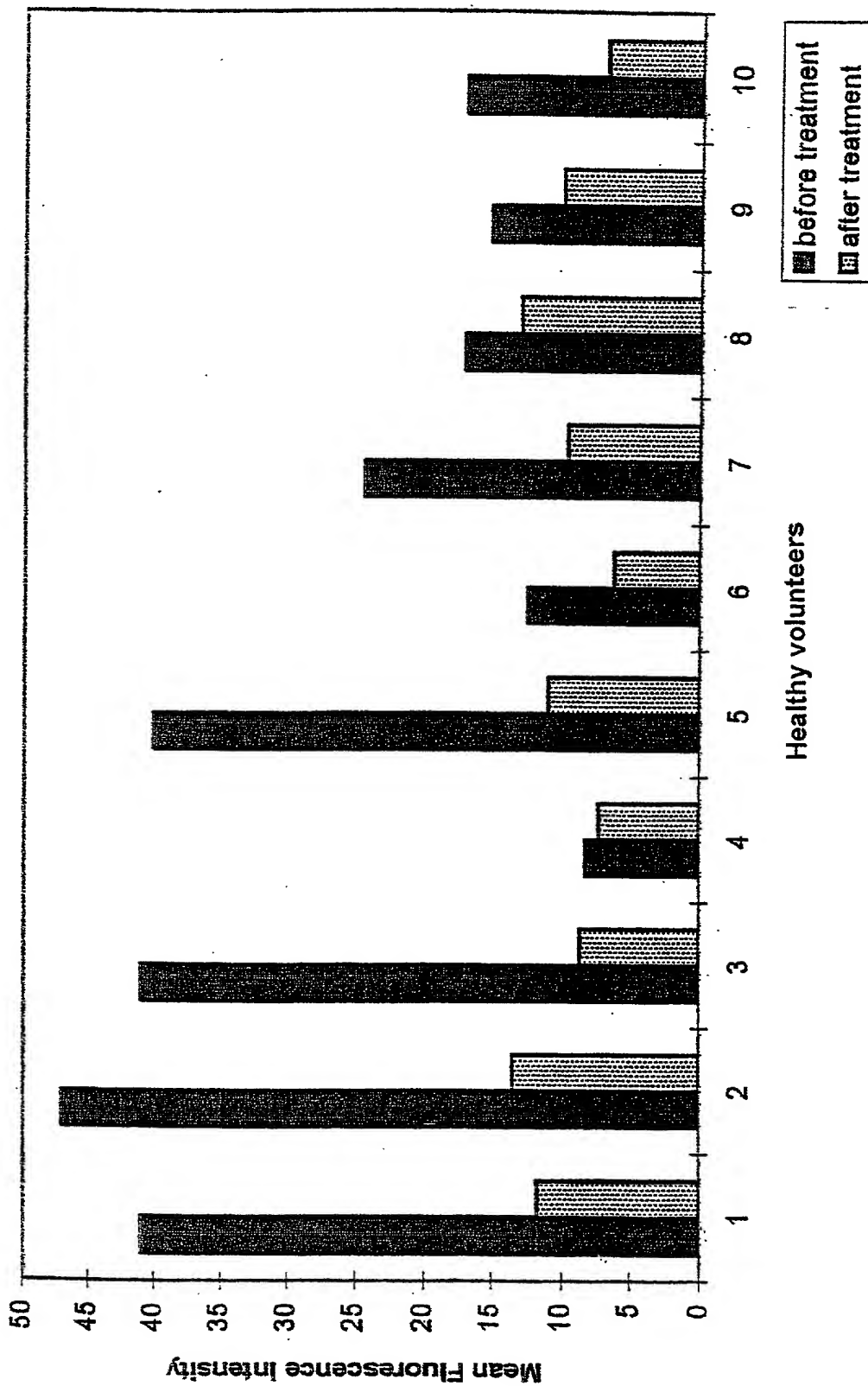


FIG. 3



PRO 10
US

Declaration, Power Of Attorney and Petition

Page 1 of 3

WE (I) the undersigned inventor(s), hereby declare(s) that:

My residence, post office address and citizenship are as stated below next to my name,

We (I) believe that we are (I am) the original, first, and joint (sole) inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled

REDUCTION OF OXIDATIVE STRESS FACTORS

the specification of which

☐ is attached hereto.

☐ was filed on _____ as

Application Serial No. _____

and amended on _____

☒ was filed as PCT international application

Number PCT/SE99/01741

on October 1, 1999,

and was amended under PCT Article 19

on _____ (if applicable).

COPY

We (I) hereby state that we (I) have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

We (I) acknowledge the duty to disclose information known to be material to the patentability of this application as defined in Section 1.56 of Title 37 Code of Federal Regulations.

We (I) hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s)

Application No.	Country	Day/Month/Year	Priority Claimed
9803334-3	Sweden	01 October 1998	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
9900371-7	Sweden	04 February, 1999	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No
			<input type="checkbox"/> Yes <input type="checkbox"/> No

We (I) hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below.

_____ (Application Number)	_____ (Filing Date)
_____ (Application Number)	_____ (Filing Date)

We (I) hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

Application Serial No.	Filing Date	Status (pending, patented, abandoned)
PCT/SE99/01741	01 October 1999	
_____	_____	_____
_____	_____	_____

And we (I) hereby appoint: Norman F. Oblon, Reg. No. 24,618; Marvin J. Spivak, Reg. No. 24,913; C. Irvin McClelland, Reg. No. 21,124; Gregory J. Maier, Reg. No. 25,599; Arthur I. Neustadt, Reg. No. 24,854; Richard D. Kelly, Reg. No. 27,757; James D. Hamilton, Reg. No. 28,421; Eckhard H. Kuesters, Reg. No. 28,870; Robert T. Pous, Reg. No. 29,099; Charles L. Gholz, Reg. No. 26,395; William E. Beaumont, Reg. No. 30,996; Jean-Paul Lavalleye, Reg. No. 31,451; Stephen G. Baxter, Reg. No. 32,884; Richard L. Treanor, Reg. No. 36,379; Steven P. Weihrouch, Reg. No. 32,829; John T. Goolkasian, Reg. No. 26,142; Richard L. Chinn, Reg. No. 34,305; Steven E. Lipman, Reg. No. 30,011; Carl E. Schlier, Reg. No. 34,426; James J. Kulbaski, Reg. No. 34,648; Richard A. Neifeld, Reg. No. 35,299; J. Derek Mason, Reg. No. 35,270; Surinder Sachar, Reg. No. 34,423; Christina M. Gadiano, Reg. No. 37,628; Jeffrey B. McIntyre, Reg. No. 36,867; William T. Enos, Reg. No. 33,128; Michael E. McCabe, Jr., Reg. No. 37,182; Bradley D. Lytle, Reg. No. 40,073; and Michael R. Casey, Reg. No. 40,294; our (my) attorneys, with full powers of substitution and revocation, to prosecute this application and to transact all business in the Patent Office connected therewith; and we (I) hereby request that all correspondence regarding this application be sent to the firm of OBLON, SPIVAK, MCCLELLAND, MAIER & NEUSTADT, P.C., whose Post Office Address is: Fourth Floor, 1755 Jefferson Davis Highway, Arlington, Virginia 22202.

We (I) declare that all statements made herein of our (my) own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1-00 Marek Naruszewicz
NAME OF FIRST SOLE INVENTOR

Marek Naruszewicz
Signature of Inventor

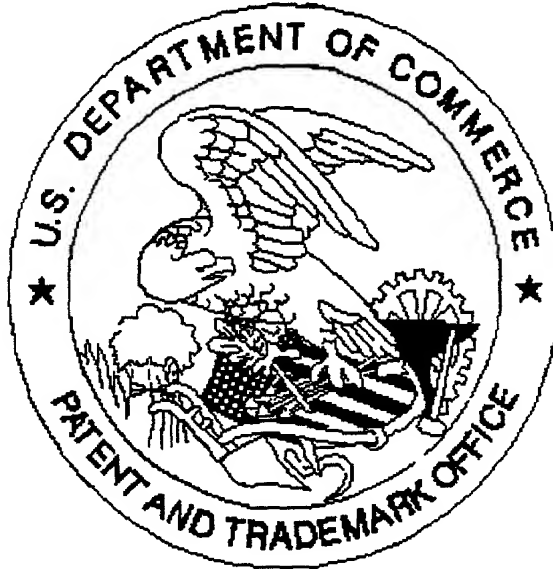
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